Mutation Analysis of Exons 10 and 17a of CFTR Gene in Patients with Cystic Fibrosis in Kermanshah Province, Western Iran

Abbas Sahami 1, Reza Alibakhshi 2*, Keyghobad Ghadiri 3, Hamid Sadeghi 1

1- Department of Biology, Jahrom Branch, Islamic Azad University, Jahrom, Iran
2- Nano Drug Delivery Research Centre, Kermanshah University of Medical Sciences, Kermanshah, Iran
3- Nosocomial Infectious Disease Research Center, Kermanshah University of Medical Sciences, Kermanshah, Iran

Abstract

Background: Cystic fibrosis (CF) is the most common genetic disorder with autosomal recessive inheritance among Caucasian populations. So far, more than 1950 different mutations were identified in cystic fibrosis transmembrane conductance regulator (CFTR) gene. CFTR gene has 27 exons. The type and distribution of mutations vary widely among different countries and/or ethnic groups. Therefore, a comprehensive analysis was performed on exon10 and exon17a of CFTR gene in CF patients in the Kermanshah province, western Iran.

Methods: We tested 27 patients admitted to the medical genetics laboratory of Kermanshah University of Medical Sciences. The patients were from different cities of Kermanshah province. All the patients had the clinical signals and two positive sweat tests. After filling agreement forms and questionnaire, the peripheral blood sampling and DNA extraction were done. DNA samples were extracted. PCR and sequencing special PCR were done. Finally analysis of the results with DNA sequencing analysis version 5.2 software was performed.

Results: CFTR mutations analysis identified 4 different mutations in our CF patients. The disease-causing mutations were p.F508del (∆F508) (14.81%), p.S466X (1.85%), and p.T1036I (1.85%). M470V polymorphism with frequency of 74.1% was found in 23 patients (17 homozygous and 6 heterozygous).

Conclusion: Three disease-causing mutations in CF patients in the present study account for approximately 18.51% of mutations. The frequency of p.F508del, the most common mutation was 16−18.1% in Iranian population. The results of the present study can be applied for genetic counseling, population screening and prenatal diagnosis.

Keywords: ∆F508, Cystic fibrosis, Direct sequencing, Iran, Kermanshah, M470V, S466X, T1036I.


Introduction

Cystic fibrosis (CF; MIM# 219700) is the most common lethal autosomal recessive disorder in Caucasian population; it affects 1 in 2500 newborns among Caucasians (1, 2). Cystic fibrosis (CF) is caused by mutations in cystic fibrosis transmembrane conductance regulator (CFTR) gene, which encodes a cAMP-dependent chloride channel that is found at the apical membrane of epithelial cells, including respiratory epithelia and submucosal glands, exocrine pancreas, liver, sweat ducts, and the reproductive tract. CFTR is a member of the ATP-binding cassette
(ABC) membrane transporter superfamily that includes proteins such as the multiple drug resistance protein (MDR) and bacterial periplasmic permeases (1, 3). CFTR gene is localized on the long arm of chromosome 7 (7q21-34), spanning approximately 190 kb of genomic DNA. The gene consists of 27 exons and encodes a mature mRNA transcript of 6.5 kb that is translated into a 1480 amino acid protein (3). To date, more than 1950 mutations (4) have been identified in cystic fibrosis transmembrane conductance regulator (CFTR) gene (4). Many of these mutations are rare in the population and based on the reports only a few affected individuals. In addition to ∆F508 mutation, these mutations vary greatly in their frequency and distribution, but most of them are very rare. Only four mutations (p.G542X, p.N1303K, p.G551D, and p.W1282X) have overall frequencies greater than 1% (5). CF remains as a life-threatening autosomal recessive condition affecting Caucasians (5). In the US, each year 1000 new cases are diagnosed. The CF incidence is estimated to be 1.2500 to 1.90,000 live births (1). In the case of CFTR, different patterns of migration and settlement have led to worldwide variation of the mutations CF is rare and less documented among Asians (1, 2). The exact incidence of CF is not known but the predictions show the variation of 1.10,000 to 1.350,000 of estimated frequencies. Minimal information is available concerning the prevalence of CF in Iran. Many children with CF in these populations probably remain undiagnosed due to lack of clinical suspicion and proper diagnostic facilities. The objective of this study was to determine the presence of CFTR gene mutations (exons 10 and 17a) among 27 CF patients in the population of Kermanshah province with respiratory and gastrointestinal manifestations similar to those reported in CF patients.

The importance of study: Adequate and accurate information of CF mutations in particular populations provides information for CF prevention programs applicable via heterozygote screening and prenatal diagnosis (6). The identification of mutations and their frequencies are thus critically important for designing gene probes for effective diagnosis of CF in a given population. Further, different mutations are associated with varying severity and prognosis management (7). Among different provinces of Iran, high genetic heterogeneity has been observed (8, 9). Additionally, CFTR mutations naturally reveal considerable heterogeneity. So, the molecular characterization of CF in Kermanshah province, as a diverse and mainly Kurdish populated area in Iran, seems a necessity.

Methods
This study has been done in Medical Genetics Laboratory of Kermanshah University of Medical Sciences in Iran. The study population included patients attending the pediatric outpatient clinics (pediatric chest and genetics clinics) and those who were admitted in the pediatric medicine or surgery wards with a suspected diagnosis of CF.

Patients: From 2011 to 2012, 27 unrelated families who had an affected child with CF were referred to the Medical Genetics Lab, Kermanshah University of Medical Science, Kermanshah, Iran. Peripheral blood samples of CF patients (sweat chloride >60 mmol/l diagnostic of CF and clinical symptoms characteristic of CF) were collected in EDTA tubes. Many patients had pulmonary complications and pancreatic insufficiency. The patients (27) were from many different regions of Kermanshah province. All patients were fully informed that their blood would be used for molecular investigation of CFTR gene mutations, and consent was obtained from each patient or his/ her guardians. Genomic DNA was collected from peripheral whole blood using the "salting out" procedure (12).

Sweat test: Sweat samples were gathered by the Macroduct sweat collection system (Wescor, Logan, UT). Localized sweating was stimulated by the iontophoresis of pilocarpine into the skin of the flexor surface of the forearm or thigh. Sweat was then collected in micro bore tubing (Macroduct), the amount of sweat was quantitated, and

<table>
<thead>
<tr>
<th>Cystic fibrosis primer name</th>
<th>Exon amplified</th>
<th>Tm</th>
<th>Primer sequence (5’-3’)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CF10-F</td>
<td>10</td>
<td>57</td>
<td>5’-TTGGAGGCAAGTGAATCC- 3’</td>
</tr>
<tr>
<td>CF10-R</td>
<td>10</td>
<td>57</td>
<td>5’-CGATTGAATATGGAGCC- 3’</td>
</tr>
<tr>
<td>CF17a-F</td>
<td>17a</td>
<td>55</td>
<td>5’-TAAATCACTGACACACATTTTGCCA- 3’</td>
</tr>
<tr>
<td>CF17a-R</td>
<td>17a</td>
<td>55</td>
<td>5’-GTACACCAACTGTGGTAAGA- 3’</td>
</tr>
</tbody>
</table>
the sample was then analyzed for chloride concentration, using the standard ISE method.

**Mutation analysis:** PCR amplification of exon10 and 17a of CFTR gene was performed using GeneAmp PCR System 9700 (Applied Biosystems, USA). The primers used in this study were synthesized by Operon (Metabion, Germany). The sequences of primers, listed in 5’-3’ direction are shown in the (Table 1). All samples were sequenced in both the forward and reverse direction using the same primers used in the PCR reactions. PCR products were purified using QIAquick PCR purification kit. For sequencing analysis, samples were analyzed by direct sequencing of exon10 and 17a of CFTR gene. The sequences were analyzed using the ABI PRISM 3100 Genetic Analyzer (Applied Biosystems, USA).

![Figure 1. Nucleotide alterations identified by sequencing](http://www.jri.ir)
17a of CFTR gene and their flanking introns in an ABI-3130 DNA analyzer (Applied Biosystems, USA). The sequences were compared with the wild-type CFTR nucleotide sequence using Seqscape software (Applied Biosystems). All identified sequence changes were confirmed by direct DNA sequencing in the reverse direction.

Results

Clinical profile and laboratory findings: A total of 27 unrelated patients (13 males and 14 females) aged two months to 19 years originating from Kermanshah province were analyzed in this study. 66.7% of the patients were from consanguineous marriage (mostly between first cousins).

In our investigation, nearly 80% of the patients had malnutrition, abnormal stools and failure to thrive, 22 cases had been diagnosed with meconium ileus and intestinal obstruction. All patients had either persistent or acute symptoms of respiratory. There are numerous studies which have reported similar results.

Genotype analysis: Mutation screening of CFTR gene in 54 alleles by sequencing reaction for all common mutations (exon 10 and exon 17a) showed that 10 alleles were ΔF508 (14.81%), S466X (1.85%) and T1036I (1.85 %) and also showed 40 alleles (74.1%) with M470V polymorphism. Five patients were heterozygous for ΔF508, one patient was homozygous for ΔF508 and one patient was a compound heterozygous (ΔF508/S466X) (Table 2, Figure 1). All had respiratory difficulties and failure to thrive. In this research, M470V polymorphism frequency of 74.1% was observed in 23 patients. M470V polymorphism was 63% in homozygote genotype patients and it was 11.1% for the heterozygote genotypes (Figure1).

Discussion

Different ethnic groups and tribes live in Iran and ethnic/genetic heterogeneity has resulted in a high number of different mutations that account for CF. Comparison between different provinces showed that the mutation spectrum differs substantially in types and frequencies (8–10).

To evaluate and analyze our results, there is a need for some general facts about the ethnicity of the people who were investigated in this study.

<table>
<thead>
<tr>
<th>Gene Location</th>
<th>Nucleotide change</th>
<th>Mutation type</th>
<th>No. of Patients</th>
<th>Global distribution</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exon 10</td>
<td>Deletion of CTT from 16533</td>
<td>p.F508del</td>
<td>1</td>
<td>6 Global</td>
</tr>
<tr>
<td>Exon 10</td>
<td>C to G at 1529</td>
<td>p.S466X</td>
<td>-</td>
<td>1 Germany-Iran</td>
</tr>
<tr>
<td>Exon17a</td>
<td>C to T at 3239</td>
<td>p.T1036I</td>
<td>-</td>
<td>1 Iran</td>
</tr>
</tbody>
</table>

Table 2. CFTR gene mutations identified as a result of the study (exon 10 and exon 17a)

<table>
<thead>
<tr>
<th>Region or Country</th>
<th>Frequency of CF alleles (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>F508del</td>
</tr>
<tr>
<td>This study</td>
<td>14.81</td>
</tr>
<tr>
<td>Lebanon</td>
<td>34-37</td>
</tr>
<tr>
<td>Palestine</td>
<td>23.5</td>
</tr>
<tr>
<td>Jordan</td>
<td>7.4-12</td>
</tr>
<tr>
<td>Syria</td>
<td>1 patient</td>
</tr>
<tr>
<td>Saudi Arabia, United Arab Emirates, Oman, Qatar, Kuwait</td>
<td>12</td>
</tr>
<tr>
<td>Saudi Arabia</td>
<td>13-15</td>
</tr>
<tr>
<td>Algeria</td>
<td>16.7</td>
</tr>
<tr>
<td>Bahrain</td>
<td>7.7</td>
</tr>
<tr>
<td>Turkey</td>
<td>24-27</td>
</tr>
<tr>
<td>Pakistan</td>
<td>17-56</td>
</tr>
<tr>
<td>Tunisia</td>
<td>18</td>
</tr>
<tr>
<td>Indian</td>
<td>19-40</td>
</tr>
</tbody>
</table>

* Some reports about this mutation (S466X) in Italy's northeast, France's northwest, Turkey, Greece and India
This is the first time that such a study is done for Kurdish people in Islamic Republic of Iran. The Kurdish people or Kurds are part of the Iranian people native to the Middle East, which includes adjacent parts of Iran, Iraq, Syria, and Turkey (11).

Kermanshah province has the most diverse Kurdish population. It even extends to some other neighboring ethnics such as Lorr, Turk and Persians. This province is located in neighbor of the provinces: Hamadan, Lorestan, Ilam and Kurdistan. The west part of this province is along Iraqi borders. Due to these unique geographical, ethnic and lingual properties of this region, "screening" of Kurdish people for this illness is quite important.

ΔF508 mutations: Only ten disease-causing mutations were identified in this study. ΔF508 was present on 8 (14.81%) of all 54 CF chromosomes analyzed (11.1% heterozygote and 3.7% homozygote). Although the frequency of ΔF508 among these patients was lower (14.81%) than its frequency in European countries (Table 3), it was still the most frequent mutation among those reported to date in Iran (8–10). Previous studies representing overall Iranian CF patients reported a frequency of 16–18.1% for this mutation (9, 10). ΔF508 is the most common mutation in the world and this mutation accounts for ~70% of worldwide (7) mutated alleles of CF chromosomes but with great variation of frequency from 100% (Faroe Islands) (11) to 18% (Tunisia). Most of the mentioned chromosomes share the same but rare haplotype for a one occurrence mutation in a normal population. ΔF508 mutation distribution shows a decreasing frequency in Europe. In the case of Middle East, ethnic background and consanguinity account for the variation. The preliminary reports on the mutation spectrum in Iranian CF children indicate that p.F508del is the most common mutation, accounting for about 16–25% of the mutations. Identification of the mutations is of great importance for further evaluation, patient counseling and prenatal diagnosis.

Table 4. Comparison of the frequency of common CFTR mutations, ΔF508 and S466X in Iran

<table>
<thead>
<tr>
<th>Frequency</th>
<th>ΔF508 (%)</th>
<th>Frequency</th>
<th>S466X (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alibakhshi et al. (2006)</td>
<td>18.1</td>
<td>5.8</td>
<td></td>
</tr>
<tr>
<td>Elahi et al. (2006)</td>
<td>16</td>
<td>1.66</td>
<td></td>
</tr>
<tr>
<td>The present study (2012)</td>
<td>14.81</td>
<td>1.85</td>
<td></td>
</tr>
</tbody>
</table>

The frequency of this mutation in western Asia and north of Africa is less than Europe. These frequencies were compared in table 3 (13–27).

ΔF508 mutation frequency is decreasing from northwest of Europe to its southeast, in such a way that this frequency in Denmark is 88% (28), in England is 74% (29), in Germany is 71% (30), in Swiss 71%, in France 69% (30), in Bulgaria 60%, in Spain 53% (31), in Sweden 52% (30), in Greece 52% (32) and in Italy 45% (30). Because of decreasing mutation frequency from northwest to southeast of Europe, it is obvious that it is about 24–27% in Turkey (33) and also it is clear that Iran's mutation frequency is similar to Turkey's.

**S466X mutation:** This mutation is the most widespread one among Iran's proportional mutations and in contrast to other reported mutations has remarkable percentage in the country. In previous studies, the percentage of homozygote of this mutation has reached to 5.8%. In the present study, this mutation is observed for the first time in Iran as compound heterozygous and this should be noted that heterozygote genotype along with mutation of ΔF508 (1.85%) were found in one of the patients. This patient was not born out of consanguinity marriage and this patient showed the symbols of malnutrition, respiratory problems and meconium ileus. In previous studies, in the common wealth of independent states (CIS), the mutation of S466X with ΔF508 in the form of compound heterozygous was determined. This mutation is usually connected with pancreas deficiency (Table 4). This deficiency was observed in our study with the same condition. S466X mutation with homozygote genotype was previously reported in Tehran, Khorasan, Hamedan and Markazi provinces. It seems that this mutation exists among many families in different regions of Iran and it seems relatively ubiquesties in Iran. Nonetheless, this mutation is very rare worldwide; for example, the frequency of 0.5% has been reported in Serbia and Montenegro. There are also some reports about this mutation in Italy’s northeast, France’s northwest, Turkey, Greece and India (16, 17, 30, 32–34).

**T1036I mutations:** In exon 17a, T1036I mutation is reported as a heterozygote for the second time in the world. This mutation was reported for the first time in 2006, in Iran (8). This mutation may be connected to low frequency of studied patients and the common tribal marriages in Iran and west
Asian cultures. However, this mutation has been reported once and was originated from Iran. The frequency of T1036I mutation (C to T at 3239) was 1.85% in this study and it was involved with just one patient (Figure 1 and Table 2). The patients had the disorders of respiratory-digestive and meconium ileus and the parents of the patient had no genetic relationship (1).

**M470V polymorphism:** Polymorphism of M470V is a silent polymorphism which changes one nucleotide and converts the methionine amino acid to valine in the place of 42376223. This polymorphism has little or no effect on the protein function. However, M470V is not strongly neutral because a study showed that this polymorphism leads to degradation of CL transitions through the cell’s membrane. It has also been showed that whenever M470V polymorphism comes along with another polymorphism mutation that leads to less activation of CFTR protein function (8, 35). This polymorphism is also effective in exon 9 process and causes genetic faulting. There are some reports worldwide which show M470V polymorphism with T5 and T7 allele (35) from Tn polymorph placement cause illness in combination (T repetitions are studied in CF patients and men’s infertility) (36). Some reports also evaluated the potential connection of men’s prostate cancer with this polymorphism (37, 38). The effectiveness of this allele in the illness phenotype is not completely clear yet and it deserves more investigations.

The incidence of cystic fibrosis is high in case of Europeans. Medical community’s lack of knowledge of the disease, poor access to medical facilities and health care for CF patients, confounding diagnosis, a high infant mortality rate, and low life expectancy in general could account for the unexplained low CF incidence in some countries with a large proportion of Caucasians of European origin. In some cases, intermediate sweat chloride levels may exist in some patients, and this may lead to misdiagnosis of them.

The CF frequency might be higher for some isolated populations due to consanguinity. In case of Turkey and Saudi Arabia, the same consideration can be mentioned. Hence, CF can be regarded as a prevalent illness even outside Europe and United State which may have different mutation spectrums. In this regard, ascertainment of CFTR mutation carrier frequencies and the CF incidence are necessary among Iranian populations.

**Conclusion**

Frequency of ΔF508, S466X and T1036I mutations in this study are quite comparable to similar studies in Iran and neighboring regions. The remaining mutations and their frequency identification call for a need to design some specific tests to achieve more precision in clinical diagnosis. Moreover determination of the full spectrum of mutations in CFTR gene in Kermanshah province entails the analysis of remaining exons in CF patients. Establishment of CF prevention programs by carrier screening and prenatal diagnosis seems to be an essential program in genetic counseling and prenatal care of carrier families.

**Acknowledgement**

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**Conflict of Interest**

The authors declare no conflict of interest.

**References**


